

Contributions to the Entomofaunal Inventory of Finca Las Piedras (Madre De Dios, Peru), Part 1: Cleridae (Coleoptera)

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**CONTRIBUTIONS TO THE ENTOMOFAUNAL INVENTORY OF
FINCA LAS PIEDRAS (MADRE DE DIOS, PERU), PART 1:
CLERIDAE (COLEOPTERA)**

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Abstract.—A list of checkered beetles (Coleoptera: Cleridae) collected at Finca Las Piedras (Madre Dios, Peru) is presented. Eight new country records for Peru, field observations and taxonomic notes are presented, and distributions are updated for all species herein. The species *Phyllobaenus cylindricollis* (Gorham, 1886), formerly not known from South America, is supplemented with data from additional geographic regions. Taxonomic problems involving *Phyllobaenus cylindricollis* (Gorham, 1886), the genus *Plocamocera*, *Enoclerus bipartitus* (Schenkling, 1915), *E. flavibasis* Chapin, 1927 (**status restored**) and *E. dichrous* Chapin, 1927 are reviewed and include the assessment of type specimens.

Key words: Brazil Nut Corridor, Manu-Alto Purus-Amarakaeri complex, Tambopata-Bahuaja Sonene-Madidi complex, entomofauna, *Phyllobaenus*

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Located near the regional capital of Puerto Maldonado in the ‘Brazil Nut Corridor’ of Madre de Dios, Peru, the habitats of the Finca Las Piedras (FLP; Fig. 1) field station and its surroundings link the flora and fauna of nearby critical protected areas of high biodiversity, including the Manu-Alto Purus-Amarakaeri and Tambopata-Bahuaja



Fig. 1. Map of southern Peru featuring the location of Finca Las Piedras; (inset) map of South America indicating (square) the region of Peru.

Sonene-Madidi complexes. To date, over 2000 species of insects have been recorded from FLP, with a significant bias towards Lepidoptera, which is the main taxonomic focus of the Alliance for a Sustainable Amazon, a research-focused nonprofit organization that manages the field site. Recent efforts are expanding this focus to include other orders of insects, including the largest lineage of described life, Coleoptera. Prior to these efforts, there are no species of checkered beetles (Coleoptera: Cleridae) that have been recorded on the property. Burke and Chaboo (2015) produced the first checklist of Peruvian Cleridae, which was subsequently updated by Juárez-Noé and González-Coronado (2024).

A collecting expedition in January 2022 was endeavored with the goal of significantly augmenting the known entomofauna of FLP.

The entomologists present had broader taxonomic training and were able to sample several groups across the property. Since the three entomologist expeditioners all studied families with preferences for dead and dying trees (Calopterygidae, Cleridae, Nitidulidae), treefalls were often favored collecting sites.

The numerous fallen trees at the site of a large Brazil nut treefall (Figs. 2–5; *Bertholletia excelsa* Humb. and Bonpl. (Lecythidaceae)) proved to be the most productive longhorn-beetle (Cerambycidae) collecting of the expedition. During initial inspections of the treefall (Figs. 2, 4), two species of checkered beetles were collected in large series. For this reason, this site became the primary location for subsequent light collecting and long-term Malaise trapping at the edge of the clearing formed by the compound treefall (because



Figs. 2–5. 2, *Bertholletia excelsa* Humb. and Bonpl. (Lecythidaceae) treefall, trunk side with GSP and SMB. 3, *Bertholletia excelsa* Humb. and Bonpl. (Lecythidaceae) treefall, canopy side and clearing. 4, Evening inspection of the compound treefall site (i.e., canopy side) with GSP discovering the presence of many adult *Plocamocera*. 5, Malaise trap along edge of compound treefall.

the large Brazil nut tree subsequently felled numerous other species of smaller trees and injured many more) (Fig. 5).

This work represents the first in a series of papers supplementing the entomofaunal inventory of FLP for the project “Species Diversity, Abundance, Distribution and Ecology in Southeastern Peru”, which is a long term, entomofauna-focused biological inventory led by the Alliance for a

Sustainable Amazon. The present work additionally supplements the recent checklist of Peruvian Cleridae (Juárez-Noé and González-Coronado 2024), which accounts for 161 identified species. The present work increases that number to 169.

MATERIALS AND METHODS

Finca Las Piedras (FLP) is the research and education center of the Alliance for a

Sustainable Amazon (<https://www.sustainableamazon.org/finca-las-piedras>). January 7–20, 2022, the authors (JML, GSP, SMB) collected east of the small town of Monterrey, including at FLP and in a forest concession for the sustainable harvest of Brazil nuts (*Bertholletia excelsa* Humb. and Bonpl. (Lecythidaceae)) ca. 20 km to the east (Madre de Dios, Peru), employing sweep-netting, aerial netting, beat-sheeting, hand-collecting at treefalls and dead or dying standing trees, fermentation traps and mercury vapor and black lights. A Malaise trap was set on January 16, 2022, at the perimeter of a compound treefall (Fig. 5) at FLP where signs of infestation were observed. The primary treefall was a large Brazil nut tree (Fig. 2; trunk end), which caused secondary treefall of several smaller trees of varying taxa (Figs. 3–5). The Malaise trap was serviced through August 1, 2022, by ZEA. Labels read “treefall” Malaise trap due to the variety of fallen trees in the immediate clearing. Collections made by hand directly from the Brazil nut tree are indicated.

Specimens were photographed with a Nikon Digital Sight DS-Fi2 imaging system mounted on a Nikon SMZ18 stereomicroscope. Full image stacks were combined using Helicon Focus 6 (www.heliconsoft.com/heliconsoftproducts/helicon-focus) and edited using Adobe Photoshop Elements 12 Editor (<https://www.adobe.com/products/photoshop-elements.html>), which was additionally used to assemble figure plates. Unless stated otherwise, figured specimens of Cleridae are exemplars from the JMLC. Robert Finn (National Identification Services, USDA-APHIS-PPQ-PEIP) provided photographs of the holotypes of *Enoclerus dichrous* Chapin, 1927 and *E. flavibasis* Chapin, 1927, both of which were examined physically by JML during a prior visit to the National Museum of Natural History (Smithsonian Institution, Washington, D.C.). The holotype of *Enoclerus bipartitus* (Schenkling, 1915)

was examined and photographed by JML in the Senckenberg Deutsches Entomologisches Institut (Müncheberg, Germany).

For each genus (and species therein), identifications were checked using primary literature, and label data were compared to the literature to expand the known distributions for each species. Country records were elucidated from specimen label data and literature. Locality data representing distributional records are presented below under “Specimens Examined.” These label data are standardized to present country, region, city, additional notes, standardized date (in the form of day-month-year as 1-II-1903), collector, depository and number of specimens, in that order. Otherwise, these data are presented verbatim.

Voucher specimens from Peru are deposited in the JMLC (John Leavengood, private collection, Brandon, Florida, USA). Duplicate specimens will be deposited in the Museo de Historia Natural of the Universidad Nacional Mayor de San Marcos (UNMSM; Lima, Peru). Additional specimens of *Phyllobaenus cylindricollis* (Gorham) were examined for additional distributional data.

Natural history museum and private collection codens are as follows:

ACMT*	American Coleoptera Museum (James E. Wappes), San Antonio, Texas
BMNH	The Natural History Museum, London, United Kingdom
CASC	California Academy of Sciences, San Francisco, California
CMNH	Carnegie Museum of Natural History, Pittsburgh, Pennsylvania
EMUS	Utah State University, Logan, Utah
FMNH	Field Museum of Natural History, Chicago, Illinois
FSCA	Florida State Collection of Arthropods, Gainesville, Florida
JMLC	John M. Leavengood, Jr., private collection

JPHC	Jeffrey P. Huether, private collection
MNHN	Muséum National d'Histoire Naturelle, Paris, France
NMNH	National Museum of Natural History, Washington, D. C.
RDCC	Ron D. Cave, private collection
RFMC	Roy F. Morris, private collection
RHTC	Robert H. Turnbow, private collection
SDEI	Senckenberg Deutsches Entomologisches Institut, Müncheberg, Germany
TAMU	Texas A and M University, College Station, Texas
UNMSM	Museo de Historia Natural, Universidad Nacional Mayor de San Marcos, Lima, Peru

*Also published as the JEWG, a large portion of this collection was accessioned by the FSCA as donations

RESULTS

The taxonomy presented reflects Gimmel et al. (2019) for subfamily composition and Bartlett (2021) for tribal composition within the subfamily Clerinae. Since the most recent revisionary work on New World Cleridae follows Opitz (2010b), subfamily classification still presents the former status of subfamily Hydnocerinae. Subheadings include the former status of the respective taxa reviewed herein. In some cases, Corporaal (1950) includes distributions not listed in Opitz' revisions of korynetine genera. Since Opitz historically omits the majority of literature on the genera being revised and includes little to no taxonomic discussions, it is unclear whether many of these inconsistencies resulted from erroneous omissions, if the specimens in question represent species subsequently described by Opitz or were discovered to be misidentifications.

Subfamily Korynetinae Laporte, 1836

Ellipotoma tenuiformis Spinola, 1844b (formerly subfamily Epiphloeinae) (Fig. 6)

Distribution.—Brazil, Colombia, Costa Rica, Ecuador, Guyana, Honduras, Panama and Peru (Opitz 2006). In Peru it is known from Amazonas, Huánuco (Opitz 2006), and now Madre de Dios.

Specimens Examined.—PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 3-VII-1-VIII-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1).

Epiphloeus erwini Opitz, 2010c (formerly subfamily Epiphloeinae)

Distribution.—Ecuador (Opitz 2010c). Peru is a **new country record**.

Specimens Examined.—PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 26-II-7-V-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1).

Ichneia marginella (Klug, 1842) (formerly subfamily Epiphloeinae) (Figs. 7–8)

Distribution.—Belize, Bolivia, Brazil, Colombia, Costa Rica, Ecuador, French Guiana, Guatemala, Guyana, Honduras, Mexico, Nicaragua, Panama, Paraguay, Peru, Suriname, Trinidad and Venezuela (Opitz 2010a). In Peru it is known from Huánuco, Loreto (Opitz 2010a), and now Madre de Dios.

Specimens Examined.—PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 26-II-7-V-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W,



Figs. 6–12. 6, *Elliptotoma tenuiformis* Spinola, 1844. 7–8, *Ichnea marginella* (Klug, 1842). 9–10, *Iontoclerus sericeus* (Klug, 1842). 11–12, *Pyticara duponti* Spinola, 1844.

treefall Malaise trap, 3-VII-1-VIII-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 2).

***Iontoclerus sericeus* (Klug, 1842)** (formerly subfamily Epiphloeinae)
(Figs. 9–10)

Distribution.—Bolivia, Brazil, Colombia, Ecuador, French Guiana and Peru (Opitz 2008). In Peru it is known from San Martín (Hualaga), Huanuco, and Madre de Dios (Juárez-Noé and González-Coronado 2024).

Specimens Examined.—PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise

trap, 20-I-26-II-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1).

***Megaphloeus mucroreus* (Klug, 1842)** (formerly subfamily Epiphloeinae)

Distribution.—Argentina, Brazil, Colombia, French Guiana, Guyana, Paraguay, Peru and Venezuela (Opitz 2011). In Peru it is known from Huánuco, Madre de Dios and San Martín (Opitz 2011).

Specimens Examined.—PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 20-I-26-II-2022,

Z. Escalante and JM Leavengood, Jr. (JMLC, 1).

***Megaphloeus sexplagiatus* (Kuwert, 1893)**
(formerly subfamily Epiphloeinae)

Distribution.—Belize, Brazil, Colombia, Ecuador, French Guiana, Panama, Peru, Venezuela (Opitz 2011), and Bolivia (Corporaal 1950). Opitz (2011; under *sexplagiatus*) cites Corporaal (1950), but does not include Bolivia, whereas Corporaal does. In Peru it is known from Huánuco, Madre de Dios and San Martin (Opitz 2011).

Specimens Examined.—PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 26-II-7-V-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 8); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 7-31-V-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 8); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 31-V-3-VII-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 4); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 3-VII-1-VIII-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1).

***Megaphloeus variegatus* (Klug, 1842)** (formerly subfamily Epiphloeinae)

Distribution.—Bolivia, Brazil, Colombia, Ecuador, French Guiana, Guyana, Peru and Venezuela (Opitz 2011). In Peru it is known from Huánuco, Loreto, San Martin, Junin (Satipo) (Opitz 2011), and now Madre de Dios.

Specimens Examined.—PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 3-VII-1-VIII-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1).

***Pelonium lampyroides* Spinola, 1844a**
(formerly subfamily Peloniinae Opitz)

Distribution.—Panama, Colombia (Opitz 2018) and Brazil (Corporaal 1950). Opitz (2018; under *lampyroides*) cites Corporaal (1950), but does not include Brazil, whereas Corporaal does. In Peru it is now known from Madre de Dios, **new country record**.

Specimens Examined.—PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 3-VII-1-VIII-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1).

***Plocamocera* Spinola, 1844a**
(formerly subfamily Epiphloeinae)

Remarks.—Eighteen specimens of two species of *Plocamocera* have been collected at the treefall (Figs. 4–5), inspiring the placement of the Malaise trap. Two specimens were collected within 100 feet of the treefall clearing on flowering Poaceae. Despite comparison with Opitz-identified voucher material at the FSCA (including paratypes) from his revision of the genus (Opitz 2004), confident identifications are difficult. The descriptions in the revision are insufficient, the revision lacks habitus photographs entirely, and errors in the revision are numerous (see Errata).

Errata (for Opitz 2004).—Couplet 4 refers to figure 149 in reference to antennal form (but the figure for the species in question (fig. 141) disagrees with the described form) whereas 4' refers to the slender form observed in figure 141, making it unclear how to correct this couplet [an emended couplet in Opitz (2010c) does not clarify the confusion of figure labeling in Opitz (2004)]; Figure 141 is not listed in any of the species treatments; Couplet 9', change figure references to 189 and 197 (from 196 and 158); Couplet 20', change figure reference to 185 (from 162).

***Plocamocera near minima* Opitz, 2004**

Remarks.—The present species keys to *P. minima* (Opitz 2004) due to its small size, entirely black head, the form of the antennal club and the multispinous protibia. However, the elytral pattern, antennal form and distribution all appear conspicuously different. Whereas the specimen was directly compared to Opitz-determined specimens (i.e., suggesting *P. c.f. minima*), it is herein suggested that the present species is likely an entirely different, undescribed species (hence *P. “near minima*).

Distribution.—*Plocamocera minima* is known from Panama.

Specimens Examined.—PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, at treefall, 7-20-I-2022, Colls.: S. Bybee, G.S. Powell and J. M. Leavengood, Jr. (JMLC, 10); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, on flowering Poaceae, 7-20-I-2022, Colls.: S. Bybee, G.S. Powell and J. M. Leavengood, Jr. (JMLC, 2); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 20-I-26-II-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1).

***Plocamocera taruma* Opitz, 2004**

Errata (for Opitz 2004).—The species treatment for *P. taruma* lists figures 162 (change to 160).

Distribution.—Known from Brazil (Opitz 2004). Peru is a **new country record**.

Specimens Examined.—ERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 26-II-7-V-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 2); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 3-VII-I-VIII-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1).

***Pyticara duponti* Spinola, 1844b** (formerly subfamily Enopliinae Gistel) (Figs. 11–12)

Remarks.—Opitz (2015) synonymized *P. flavicollis* Gorham, 1877 and *P. coronata* Gorham, 1877 under *P. duponti*.

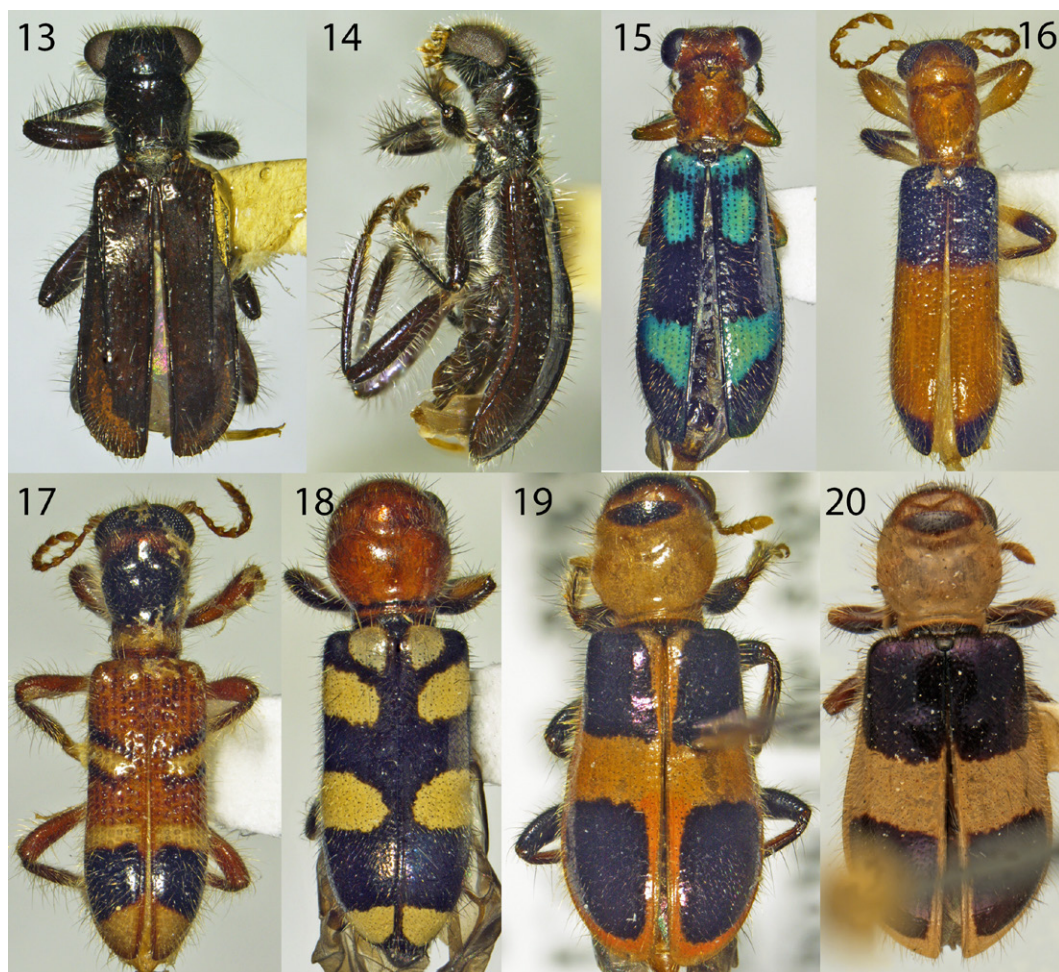
Distribution.—Brazil, Colombia, Ecuador, French Guiana (Opitz 2015), Peru (Juárez-Noé and González-Coronado 2024; as *P. flavicollis* in Burke and Chaboo 2015) and Bolivia (Corporaal 1950). Opitz (2015) erroneously omitted the type locality (Peru) of *P. flavicollis*. Opitz (2015; under *duponti*) cites Corporaal (1950), but does not include Bolivia, whereas Corporaal lists Bolivia for *Pyticara suturalis* (Schenkling, 1906) (= *P. duponti*). In Peru it is now known from Madre de Dios.

Specimens Examined.—PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 7-31-V-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1).

Subfamily Clerinae Latreille, 1802
Tribe Hydnocerini Spinola, 1844 (formerly Hydnocerinae)
Subtribe Hydnocerina Spinola, 1844
 (formerly Hydnocerini *sensu* Kolibač, 1998)

***Phyllobaenus cylindricollis* (Gorham, 1886)** (Figs. 13–14)

Remarks.—Leavengood and Garner (2014) assessed and designated/recognized eight type specimens from the BMNH and MNHN, noting two more putative syntypes in the SDEI which had not yet been examined. Döbler (1982) considered these two SDEI specimens from Bugaba and Teapa, Tabasco to be syntypes, the latter of which is decidedly incorrect considering Teapa was not included in Gorham's locality list. These SDEI specimens both have red “Syntypus” labels. But only the Bugaba specimen is truly a part of the type series. Thus, nine type specimens have been



Figs. 13–20. 13–14, *Phyllobaenus cylindricollis* (Gorham, 1886). 15, *Isolemidia pulchella* (Gorham, 1877). 16, *Axina nigrifrons* Schenkling, 1906. 17, *Priocera batesi* Opitz, 2021. 18, *Enoclerus deliciosus* (Gorham, 1876). 19, *Enoclerus dichrous* Chapin, 1927. 20, *Enoclerus dichrous* Chapin, Holotype (NMNH).

accounted. The *Teapa* non-type specimen (SDEI) mentioned by Döbler (1982) is, in fact, *cylindricollis*.

Gorham (1886) referenced “about a dozen examples” in his type series, with *P. cylindricollis* described after Gorham (1883) recognized his own error in including the then undescribed *P. cylindricollis* among specimens he considered *P. bituberculatus* (Chevrolat, 1874). Because this species was previously only known from Central America in the literature, and only questionably as far north as Guatemala and Mexico (Schenkling

1910, Wolcott 1927), additional data from other countries are presented herein in order to better explain the geographic “gap” between Costa Rica and southern Peru.

Distribution.—Costa Rica, Nicaragua and Panama (Gorham 1886, Wolcott 1927, Corporaal 1950, Barr 1975, Leavengood and Garner 2014). Mexico and Guatemala are confirmed as valid distributions. Colombia, Ecuador, Honduras and Peru are **new country records**. The distribution by Schenkling (1910), which included Mexico and Guatemala, was considered an error

(i.e., misidentification of *bituberculatus*) by Wolcott (1927) on the basis of the exclusion of those countries from Gorham's cited localities. Döbler (1982) aforementioned specimens that at least confirm the Mexico distribution. Herein we include new distributional data which include many records from southern Mexico and Guatemala. So even if Wolcott (1927) was correct in that those particular specimens reported by Schenkling were misidentified, new data reveal that both species occur in those countries, with *bituberculatus* being more commonly collected where they co-occur.

Specimens Examined.—COLOMBIA: Amazonas: PNN Amacayacu Mocagua, 150m, A. Parente (CMNH; 1). COSTA RICA: Cartago: Turrialba, Volcan Turrialba, 3-VI-1973, G. Ekis (FSCA; 1); Turrialba, 900m, VII-1925, A. Heyne (FMNH; 1); Guanacaste: 3km SE Rio Naranjo, 2-9-III, 11-20-XII, 1992, 22-25-I, 8-12-III, 21-28-III, 1-6-IV, 18-28-IV, 3-6-V, 12-14-V, 18-23-VI, 5-9-VII, 1993, F. D. Parker (EMUS; 12); Heredia: Est. Biol. La Selva, 50-150m, 10°26'N, 84°01'W, 5-8-III-2001, E. G. Riley (TAMU; 1); Estac. El Ceibo, 10km SE La Virgen, 450-550m, 7-14-IV-2003, E. G. Riley (TAMU; 1); La Selva Biol. Sta., 28-VII-1989, D. G. Furth (NMNH; 1); La Selva Biol. Sta., 12-I-1995, D. G. Furth (NMNH; 1); Limon: Escocia, on *Ingia* leaves, 15-IX-1924, F. Nevermann (NMNH; 1); Puntarenas: Las Cruces Bot Garden, 7km S San Vito, 1-17-VIII-1986, R. D. Cave (RDCC; 1); Estacion Biologica Las Cruces, 6 km S San Vito, 17-20-IV-2003 E. G. Riley (TAMU; 2); Las Alturas (San Vito), 4900' elevation, 23-24-I-1996, E. Giesbert and F. Hovore (FSCA; 1); [Province unknown]: Salvadora Farm, 19-31-XII-1930 F. Nevermann (NMNH; 1). ECUADOR: Orellana: Yasuni National Park, Onkone Gare, -76.452654, -0.658348, elev 216m, active canopy fogging, 10-I-2018, B. Garner, T. & L. Erwin, specimen Id: NHMUK014456207; Image Id: 4233-01-0030; institution code: BMNH2019-01; sequence id: BIOD06477

(BMNH, 1*). GUATEMALA: Baja Verapaz, vic. Biotopo del Quetzal, 5200' elevation, 25-V-1-VI-1989, E. Giesbert (FSCA; 1); 19-24 km north of Salama, 4500' elevation, 25-31-V-1989, E. Giesbert (FSCA; 1); HONDURAS: Copan: 6km NW San Augustin, 19-V-2002, R. Turnbow (RHTC; 1). MEXICO: Chiapas: 13 mi. northwest of Ocozocoautla, el. 3200' 16-VIII-1967, H. R. Burke and J. Hafernrik (TAMU; 1); 11 mi. n. Ocozocoautla, 19-VII-1973, Mastro and Schaffner (TAMU; 1); Lagos de Montebello, 1-VI-1987, E. Giesbert (FSCA; 2); El Sumidero Parque, 23-24-VI-1990, J. P. Huether (JPHC; 1); Veracruz: 10 mi NE El Tropico, 26-27-VI-1985, D. Heffern (NMNH; 1); Veracruz: Cordoba, A. Fenyas (CASC; 2); Atoyac, Cera Cruz, V-1907, H. H. S[mith]. (BMNH; 1). PANAMA: Chiriquí: Bugaba, Champion (BMNH; 1); Cerro Punt. Area, 13-V-1981, J. E. Wappes (ACMT; 1); Lagunas del Volcan, 5 km SW Volcan, 4220 ft, 30-VII-1999, J. B. Woolley (TAMU; 2); Cocle: Altos de Campana, 25-V-1996, R. Turnbow (RHTC; 1); Panama: 7-10 km north of El Llano, 14-22-V-1993, E. Giesbert (FSCA; 2); 7-10 km north of El Llano, 21-30-IV-1995, E. Giesbert (FSCA; 1); 10 km north of El Llano, 20-I-1996, E. Giesbert and F. Hovore (FSCA; 1); Cerro Campana, 17-V-1982, E. Giesbert (FSCA; 1); Cerro Campana, 12-III-1972, 2-IX-1972, Stockwell (NMNH; 3); Cerro Campana, 750 m, 18-VI-1996, Gillogly and Schaffner (TAMU; 1); Cerro Azul, microneadas, 20-V-1999, Morris, Wappes (RFMC; 1); San Blas Province: Nusagandi, 320m, 9°19' N, 78°55' W, 28-V-1995, A. Gillogly (TAMU; 1); Veraguas: 8 km W Santa Fe, Cerro Tute, el 3000 ft, 24-VII-1999, J. B. Woolley (TAMU; 1). PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 3-VII-1-VIII-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1). *This specimen has not been mounted or labeled, nor have labels yet been written. So, for now, the available information is presented.

Subtribe Lemidiina Kolibač, 1998

(formerly Lemidiini)

***Isolemidia pulchella* (Gorham, 1877)**

(Fig. 15)

Remarks.—Despite serious need for revision in order to reliably identify most specimens of *Isolemidia*, the single specimen presented herein matches the lectotype (illustrated in Fig. 44 of Leavengood and Garner 2014) perfectly.

Distribution.—Brazil (Leavengood and Garner 2014). Peru is a **new country record**.

Specimens Examined.—PERU: Madre de Dios: Tambopata: Concesión Vilca, nr. road marker km 21, ~21 km E of Monterrey, ca. 12.2248 S 68.9699 W, 18-I-2022, Colls.: S. Bybee, G.S. Powell and J. M. Leavengood, Jr. (JMLC, 1).

Tribe Priocerini Laporte, 1838***Axina nigrifrons* Schenkling, 1906**

(Fig. 16)

Distribution.—Bolivia, Brazil and Venezuela (Opitz 2020). Peru is a **new country record**.

Specimens Examined.—PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, 7-20-I-2022, Colls.: S. Bybee, G.S. Powell and J. M. Leavengood, Jr. (JMLC, 1).

***Priocera batesi* Opitz, 2021**

(Fig. 17)

Distribution.—Bolivia, Brazil, Colombia, Ecuador, Peru, Venezuela (Opitz 2021). In Peru it is known from Puno and Madre de Dios (Opitz 2021).

Specimens Examined.—PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, 7-20-I-2022, Colls.: S. Bybee, G.S. Powell and J. M. Leavengood, Jr. (JMLC, 1).

Tribe Clerini Latrielle, 1802***Perilypus* group sensu Bartlett, 2021*****Enoclerus deliciolus* (Gorham, 1876)**

(Fig. 18)

Distribution.—Bolivia, Brazil and Peru (Corporaal 1950). Known from Cusco (Callanga) (Peracchi 1967, Ceballos 1981), and now Madre de Dios in Peru.

Specimens Examined.—PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 20-I-26-II-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 3); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 26-II-7-V-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 7-31-V-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 2); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 31-V-3-VII-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1).

***Enoclerus dichrous* Chapin, 1927**

(Figs. 19–20, 26–27)

Remarks.—The single specimen (Fig. 19) presented herein was compared to the type (Fig. 20; NMNH). This species shares superficial similarities to the genus *Quadrophenia* Rifkind (2017), however its apical antennomeres (Fig. 27) are consistent with *Enoclerus*. In *Quadrophenia* antennomere XI is broadly emarginate internally and terminates in an acuminate apex. The present specimen also exhibits strong tibial carinations (Figs. 25–26) not addressed in Chapin's (1927) description.

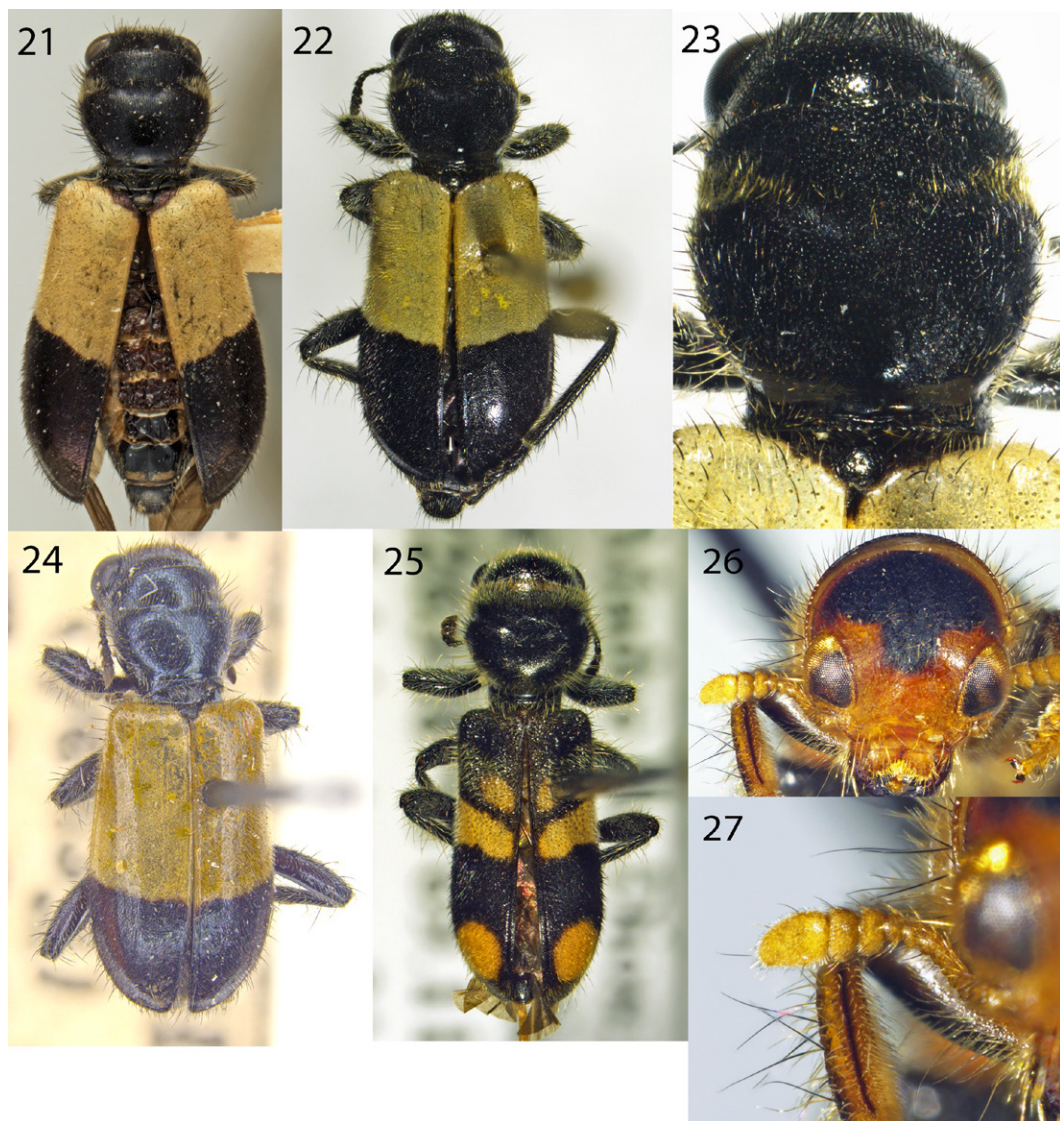
Distribution.—Bolivia (Corporaal 1950). Peru is a **new country record**.

Specimens Examined.—BOLIVIA: Mulford Biol. Expl. 1921–1922, Tumupasa, XII,

W. M. Mann, Type No. 29355 U.S.N.M. [red label], USNMENT 02019487 [QR code label] (Holotype; NMNH, 1). PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 3-VII-1-VIII-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1).

***Enoclerus flavibasis* Chapin, 1927, status restored**
(Figs. 21–23)

Remarks.— Dozens, if not hundreds, of these beetles swarmed either ventrolateral side of a felled Brazil nut tree (Figs. 2–3). Their



Figs. 21–27. 21, *Enoclerus flavibasis* Chapin, 1927, Holotype (NMNH). 22, *E. flavibasis* (JMLC). 23, Pronotum and baso-sutural region of elytra of *E. flavibasis* (JMLC). 24, *Enoclerus bipartitus* (Schenkling, 1915), holotype (SDEI). 25, *E. lugubris* (Erichson, 1847). 26, *E. dichrous* head and protibial carina, frontal view. 27, *E. dichrous* antennal club.

presumed tree-infesting prey were not discovered. Opitz (2024) placed *Enoclerus bipartitus* (Schenkling, 1915), although misspelled as “*bipartitus*”, in synonymy with *E. flavibasis*. This nomenclatural act was executed with the same broad justification as numerous other synonymies (presented in list form): “I have examined specimens of all known South American species of *Enoclerus*, including many types, and I find that on the basis of male genitalia various nominal species need to be placed in synonymy (Opitz 2024).” There were no accompanying figures, descriptive notes, specific types, nor examined specimens cited, following the practices of Barr (2018, written by Opitz).

However, the type specimen of *E. bipartitus* (Fig. 24; SDEI) has the apical one-third of the elytra black (otherwise entirely yellow-green), has the baso-sutural region of the elytra all yellow-green, and lacks distinct pale patches of lateral pronotal setae (or sometimes present, but not dense); whereas the type specimen of *flavibasis* (Fig. 21) and the fifty-five specimens presented herein all consistently differ in having just less than half of the elytra black (with minimal variation; Fig. 22), also consistently with the baso-sutural region around the scutellum black (Fig. 23), some specimens also with small black elytral humeral maculae, and moderate to strong distinct pale patches of lateral pronotal setae (Fig. 23). Although species of *Enoclerus* may exhibit intraspecific variation with respect to patches of pronotal setae (and their weak to distinct presence) and elytral coloration, the variation of the reported *flavibasis* specimens forms a distinct expression range that does not allow for the conspecific inclusion of *bipartitus*. Given the lack of evidence presented by Opitz (2024) and the consistent differences between the *flavibasis* type and specimens presented herein in comparison to the type of *bipartitus*, we hereby restore the status of *Enoclerus flavibasis*.

Distribution.—Bolivia (Corporaal 1950). Peru is a **new country record**. *Enoclerus bipartitus* is known from Ecuador (Corporaal 1950).

Specimens Examined.—BOLIVIA: Mulford Biological Exploration 1921–1922, Rurrenabaque Rio Beni, XI-1921, W. M. Mann, Type No. 29357 U.S.N.M. [red label], USNMMENT 02019488 [QR code label] (*flavibasis* Holotype; NMNH, 1). PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, at *Bertholletia excelsa* treefall, 7-20-I-2022, Colls.: S. Bybee, G.S. Powell and J. M. Leavengood, Jr. (JMLC, 11); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 16-20-I-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 4); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 20-I-26-II-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 3); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 26-II-7-V-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 7); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 7-31-V-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 8); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 3-VII-1-VIII-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 22).

Enoclerus lugubris (Erichson, 1847)

(Fig. 25)

Distribution.—Bolivia, Brazil and Peru (Corporaal 1950). Known from Loreto (Peracchi 1967), and now Madre de Dios in Peru.

Specimens Examined.—PERU: Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 16-20-I-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1); Madre de Dios: Tambopata: Monterrey, Finca las Piedras, vic. 12.226348° S 69.112599° W, treefall Malaise trap, 31-V-3-VII-2022, Z. Escalante and JM Leavengood, Jr. (JMLC, 1).

DISCUSSION

The entire tribe Hydnocerini (formerly the subfamily Hydnocerinae) was correctly absent in Burke and Chaboo (2015). There have been no formally published species records for the taxon until the present work. This is largely due to the difficulty in identifying species of *Isolemidia* Gorham, 1877 and *Phyllobaenus*. A perfect example of which are two specimens of *Phyllobaenus* collected by Gino Norbil in northern Peru (Juárez-Noé and González-Coronado 2024). Unfortunately, the species-group including *P. alboasciatus* (Lucas), *P. cinctus* (Spinola), *P. femoralis* (Chevrolat), and several more similar species require a genitalia-informed revision to confidently identify any South American specimen, whether putatively described or not. That withstanding, Juárez-Noé and González-Coronado (2024) produced the first specimens of Peruvian Hydnocerini of which author JML was aware outside of his own unpublished field, museum and specimen loan work. We expect to uncover more Peruvian species in the genera *Isolemidia*, *Phyllobaenus* and *Neohydnocera* Leavengood, 2025.

The most unexpected discovery herein is the presence of *Phyllobaenus cylindricollis* in South America. *Phyllobaenus cylindricollis* is quite distinctive in the structure of its elytra, possessing a well-developed carina forming from each elytral humeral umbone, a condition only observed in one other described species *Phyllobaenus vitrinus* (Gorham, 1886), which is otherwise distinct in sharing a form more like *P. corticinus* (Gorham, 1883) (Leavengood et al. 2012).

Yet unconfirmed is “*Plocamocera* near *minima*” and at least one species of *Enoclerus* (data not presented herein). For *P. nr. minima* no new country record is registered, nor is this species being counted towards the total number of taxa in the FLP Entomofaunal Inventory. Since Malaise trap sampling has continued at Finca Las Piedras since August 1, 2022 (the final collection date of the specimen data presented herein) by author ZEA,

we hope to present additional findings in the future.

Seventeen identified species (i.e., not counting *P. nr. minima*) of Cleridae are added to the FLP Entomofaunal Inventory (which was 2,020 species as of October 2023), bringing the total to 2037. Of these records from FLP, and *Isolemidia pulchella* (caught only at “Concesión Vilca”, a SERFOR-granted concession), we present eight new country records for Peru and three additional new country records (Colombia, Ecuador, Honduras) for *Phyllobaenus cylindricollis*.

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